

Kai Zhang, Ph.D.

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APPOINTMENT

Associate Professor, University of Illinois at Urbana-Champaign (UIUC)	2021-present
Associate Department Head, Department of Biochemistry, UIUC	2021-present
Director of Graduate Studies, Department of Biochemistry, UIUC	2021-present
Assistant Professor, Department of Biochemistry, UIUC	2014-2021
Affiliate Faculty, Neuroscience Program, UIUC	
Affiliate Faculty, Cancer Center at Illinois, UIUC	
Affiliate Faculty, Center for Biophysical and Computational Biology, UIUC	
Affiliate Faculty, Beckman Institute, UIUC	
Affiliate Faculty, Chemistry-Biology Interface Training Program, UIUC	

PROFESSIONAL PREPARATION

Postdoctoral Scholar	Stanford University, Stanford, California	2009-2014
Department of Chemistry		
Research Advisor: Dr. Bianxiao Cui		

Ph.D.	University of California, Berkeley, Berkeley, California	2002-2008
Department of Chemistry		
Research Advisor: Dr. Haw Yang		

Dissertation title: Methodology Development for Single-Molecule/Particle Optical Study of Biological Systems

B.S. University of Science and Technology of China (USTC), Hefei, Anhui, PRC	1997-2002
Department of Chemical Physics	
Research Advisor: Dr. Hongfei Wang	

HONORS AND AWARDS

Scialog Fellow Research Award	<i>Research Corporation for Science Advancement</i>	2021, 2023
Inaugural "Light Lecture" Speaker	<i>University of Cincinnati</i>	2021
Scialog Fellow	<i>Research Corporation for Science Advancement</i>	2020-2023
Spotlight on Early Career Researchers	<i>Journal of Molecular Biology</i>	2017
Innovative Teaching and Learning Grant	<i>UIUC</i>	2016

American Cancer Society Postdoctoral Fellowship	<i>American Cancer Society</i>	2013
Biophysical Society Education Travel Award	<i>Biophysical Society</i>	2013
American Society for Cell Biology (ASCB) Travel Award	<i>ASCB</i>	2012
Irving Fatt/Samuel Ruben Award	<i>UC Berkeley</i>	2004
Guo Moruo Fellowship, Top Fellowship	<i>USTC</i>	2001
Award for Best Undergraduate Research	<i>USTC</i>	2001
Asian Spectra Physics Corporation Fellowship	<i>USTC</i>	2000
Legend (now Lenovo) Fellowship	<i>USTC</i>	1999
He Duohui Academician Fellowship	<i>USTC</i>	1998
Outstanding Undergraduate Award	<i>USTC</i>	1997

GRANT REVIEW PANEL

Israel Science Foundation (2023)
 Cancer Center at Illinois (CCIL) (2023)
 Paul G. Allen Frontiers Group (2021, 2022)
 Arizona Alzheimer's Disease Core Center Grant (2017)
 Research Board, OVCR in UIUC (2017, 2018, 2019, 2020)

EDITORSHIP

Journal of Molecular Biology – Associate Editor 2018-present
 Edited work: Article collection: Chemogenetics and Optogenetics
 Frontiers in Molecular Neuroscience
 - Review Editor 2015-2019
 - Associate Editor 2020-present
 CRC Methods in Signal Transduction – Editor 2021
 Edited work: Opsin-free optogenetics – Technology and Applications 2022
 Advanced Biology – Associate Editor 2022-present

RESEARCH SUPPORT

Ongoing Research Support

R01GM132438 Zhang (PI) 09/15/2019-06/30/2024
 Precise regulation of native transcription factor at the single-cell level
 The goal of this study is to develop an optogenetic approach to control the transcriptional activity of native transcription factor.
 Role: PI

Cancer Center at Illinois #9572 Zhang (PI) Nelson (CoI) 07/01/2022-06/30/2024
 Optogenetic Modulation of the Tumor Microenvironment to Improve Cancer Immunotherapy
 The goal of this study is to use optogenetic stimulation to tune the immune landscape of the tumor microenvironment.
 Role: PI

R01 MH124827 Tsai (PI), Zhang (MPI) 12/01/2020-10/31/2025

Mechanism of Gp1 mGluR-dependent translation and plasticity

The goal of this study is to determine the role of mGluR dependent translation and neuronal plasticity.

Role: PI

NSF 2121003 Cao (PI) Zhang, Cheng, Schroeder (Co-PI) 10/01/2021-09/30/2026

GCR: Synthetic Neurocomputers for Cognitive Information Processing

The goal of this study is to understand cognitive information processing through the experimental buildup of neuronal cell networks and machine learning modeling

Role: Co-PI

NSF 2243257 Luthey-Schulten (PI), Gruebele (Co-PI)

Science and Technology Center for Quantitative Cell Biology

09/15/2023-09/14/2028

The goal of this center grant is to integrate computational and experimental biology to gain quantitative insights into the structure, dynamics, interaction, and motility that define molecular and organelle functions in live cells.

Role: Co-PI

R01GM141298 Pogorelov(PI) Zhang (CoI) 09/20/2021-07/31/2025

Capturing structure and dynamics of transmembrane signaling proteins

The goal of this project is to use computational prediction and in-cell signaling to quantitatively map out the structure-function relationship of single-transmembrane receptor tyrosine kinases.

Role: Co-Investigator

Completed Research Support

American Cancer Society PF-13-030-01-DDC, Zhang (PI) 2013/01/01-2013/12/31

Cell-fate determination by light-gated MAPK and AKT signaling pathways

The goal of this study is to compare the effects of light-activated MAPK and AKT pathway on cell proliferation and differentiation.

Role: PI

R56MH118908 Wang (PI) 08/02/2019-08/01/2020

Mechanism underlying cognitive and synaptic flexibility

The goal of this study is to determine the role of adenylate cyclases 8-PI3K/Akt-GSK3 β signaling in the regulation of synaptic depotentiation reversal/suppression of memory

Role: Co-Investigator

R03NS120516 Tsai (PI) 08/01/2020-07/31/2021

Study of PAK3 in epilepsy-associated defects in synaptic plasticity

The goal of this project is to determine how nuclear and cytoplasmic PAK3 contributes to epilepsy-associated defects in synaptic plasticity.

Role: Co-Investigator

Scialog Award #27937 Zhang (PI) Levy (PI) Ross (PI) 07/01/2021-06/30/2023

Engineering enteric neuron activity to enhance antimicrobial immunity in the gut

The goal of this study is to determine the regulatory role of enteric neuronal system in antimicrobial response.

Role: PI

R01MH124992 Wang (PI) 12/01/2020-10/31/2023

Mechanism underlying cognitive and synaptic flexibility

The goal of this study is to determine the role of adenylate cyclases 8-PI3K/Akt-GSK3 β signaling in the regulation of synaptic depotentiation reversal/suppression of memory.

Role: Co-Investigator

R01 MH119149-01 Wang (PI) 04/01/2019-01/31/2024

Nonconventional role of ADCY in Gq-mediated neuronal signaling and neuroplasticity.

The goal of this study is to determine the role of adenylate cyclases (ADCY) in the development of neuroplasticity.

Role: Co-Investigator

PATENT

Glimpse: generalizable light modulated protein stabilization system, UIUC Ref. No. UIUC2019-080-01(PRO).

PUBLICATIONS (*CORRESPONDING AUTHOR)

Complete List of Published Work in My Bibliography:

<https://www.ncbi.nlm.nih.gov/myncbi/kai.zhang.9/bibliography/public/>

Peer-reviewed journal publications – independent career (UIUC)

1. Teak-Jung Oh, Bryan Gworek, Amna Mehfooz, **Kai Zhang***, “Methods for Gene Delivery and Analysis of Optogenetic Induction of Lytic Cell Death”, *Current Protocols*, 2024. DOI:10.1002/cpz1.1023.
2. Kevin Gill, Kritika Mehta, Jeremiah Heredia, Vishnu Krishnamurthy, **Kai Zhang**, Erik Procko “Multiple mechanisms of self-association of chemokine receptor CXCR4 and CCR5 demonstrated by deep mutagenesis”, *Journal of Biological Chemistry*, 2023, 299(10) 105229.
3. Fatemeh Ostadhossein, Parikshit Moitra, Maha Alafeef, Dinabandhu Sar, Shannon D’Souza, Lily Benig, Michael Nelappana, Xuedong Huang, Julio Soares, **Kai Zhang**, Dipanjan Pan, “Ensemble and single-particle level fluorescent fine-tuning of carbon dots via positional changes of amines towards the ‘supervised’ oral microbiome sensing”, *Journal of Biomedical Optics*, 2023, 28, 8, 082087.

4. Bing Bu, Zhiqi Tian, Dechang Li, **Kai Zhang**, Wei Chen, Baohua Ji, Jiajie Diao, “Double-transmembrane domain of SNAREs decelerates the fusion by increasing the protein-lipid mismatch”, *Journal of Molecular Biology*, 2023, 435, 13, 168089.
5. Ye Tian, Matthew Jellinek, Kritika Mehta, Sun Mi Seok, Shanny Hsuan Kuo, Wei Lu, Rucheng Shi, Richard Lee, Gee Lau, Jongsook Kim Kemper, **Kai Zhang**, David Ford, Bo Wang, “Membrane phospholipid remodeling modulates nonalcoholic steatohepatitis progression by regulating mitochondrial homeostasis”, *Hepatology*, 2023, 2023, 10, 2300416.
6. Huaxun Fan[†], Collin Barnes[†], Hyojeong Hwang, **Kai Zhang***, Jing Yang*, “Precise modulation of embryonic development through optogenetics”, *Genesis, The journal of Genetics and Development*, 2022, 60 (10-12), e23505. (Invited Review).
7. Qi Wang, Xiaomin Huang, Yixun Su, Guowei Yin, Shouyu Wang, Bin Yu, Hui Li, Junhua Qi, Hui Chen, Wen Zeng, **Kai Zhang**, Alexei Verkhatsky, Jianqin Niu, Chenju Yi, “Activation of Wnt/ β -catenin pathway mitigates blood-brain barrier dysfunction in Alzheimer’s disease” *Brain*, 2022, awac236. (Front Cover)
8. Kangqiang Qiu, Weiwei Zou, Hongbao Fang, Mingang Hao, Kritika Mehta, Zhiqi Tian, Jun-Lin Guan*, **Kai Zhang***, Taosheng Huang*, Jiajie Diao*, “Light-activated mitochondrial fission through optogenetic control of mitochondria-lysosome contacts,” *Nature Communications*, 2022, 13, 4303.
9. Savanna Sharum Skeeters; Tyler Camp; Huaxun Fan; **Kai Zhang***, “The expanding role of split protein complementation in opsin-free optogenetics”, *Current Opinion in Pharmacology*, 2022, 65:102236. (Invited Review)
10. Vishnu V. Krishnamurthy, Hyojeong Hwang, Jia Fu, Jing Yang,* **Kai Zhang***, “Optogenetic control of the canonical Wnt signaling pathway during *Xenopus laevis* embryonic development”, *Journal of Molecular Biology*, 2021, 433, Issue 18, 167050. (Front Cover)
11. Teak-Jung Oh[†], Huaxun Fan[†], Savanna S. Skeeters[†], **Kai Zhang*** “Steering Molecular Activity with light: Recent Advances and Perspectives”, *Advanced Biology*, 2021, 2000180.
12. Parinaz Fathi, Ayman Roslend, Kritika Mehta, Parikshit Moitra, **Kai Zhang**, Dipanjan Pan “UV-Trained and Metal-Enhanced Fluorescence of Biliverdin and Biliverdin Nanoparticles”, *Nanoscale*, 2021, 13, 4785-4798. (Inside Front Cover)
13. Qixin Chen, Hongbao Fang, Xintian Shao, Zhiqi Tian, Yuming Zhang, Huaxun Fan, **Kai Zhang***, Weijiang He*, Zijian Guo*, Jiajie Diao*, “A dual-labeling probe to track functional mitochondria-lysosome interactions in live cells”, *Nature Communications*, 2020,11, 6290.
14. Savanna S. Skeeters, Ana C. Rosu, Divyanshi, Jing Yang, **Kai Zhang*** “Comparative determination of cytotoxicity of sub-10-nm copper nanoparticles to prokaryotic and eukaryotic cells”, *ACS Applied Materials & Interfaces*, 2020,12, 45, 50203–50211.
15. Qin Wang, Huaxun Fan, Feng Li, Savanna S. Skeeters, Vishnu Krishnamurthy, Yuanquan Song*, **Kai Zhang***, Optical control of ERK and AKT signaling promotes axon regeneration and functional recovery of PNS and CNS in *Drosophila*, *eLife*, 2020, 9, e57395.

Highlighted in the School of [MCB](#).

16. Taida Huang, Yan Zhang, Zitian Wang, Hongda Chen, Nan Wang, Huaxun Fan, Zhangsen Huang, Yixun Su, Xiaomin Huang, Hui Chen, **Kai Zhang***, Chenju Yi*, "Optogenetically-controlled TrkA activity improves the regenerative capacity of hair-follicle-derived stem cells to differentiate into neurons and glia", *Advanced Biology*, 2020, 2000134.
17. Miaoling Li§, Teak-Jung Oh§, Huaxun Fan§, Jiajie Diao*, **Kai Zhang***, "Role of SNARE protein conformation in synaptic transmission: Challenges and Perspectives", *Journal of Molecular Biology*, 2020, 432, 4773-4782. (§ These authors contributed to this work equally).
18. Yixun Su, Xiaomin Huang, Zhangsen Huang, Taida Huang, Tao Li, Huaxun Fan, **Kai Zhang***, Chenju Yi*, "Early but not Delayed RAF Activation Promotes Astrocytogenesis in Mouse Neural Progenitors Resolved by Optogenetics", *Journal of Molecular Biology*, 2020, 432, 4358-4368.
19. Vishnu V. Krishnamurthy, Jia Fu, Teak-Jung Oh, John Khamo, Jing Yang*, **Kai Zhang*** "A Generalizable Optogenetic Strategy to Regulate Receptor Tyrosine Kinases during Vertebrate Embryonic Development", *Journal of Molecular Biology*, 2020, 432, 10, 3149-3158.
20. Melody Esmaeili, Shelby Blythe, **Kai Zhang**, Jing Yang, Peter Klein "Loss of Competence in early development is mediated by loss of chromatin accessibility", *Developmental Biology*, 2020, 452, 1, 20-35.
21. Tyler Camp, Kritika Mehta, Stephen Sligar*, **Kai Zhang***, "Molecular Orientation Determination in Nanodiscs at the Single-Molecule Level", *Analytical Chemistry*, 2020, 92, 2, 2229-2236.
22. Zichen Wang, Huaxun Fan, Xiao Hu, John Khamo, Jiajie Diao*, **Kai Zhang***, Taras Pogorelov* "Coaction of Electrostatic and Hydrophobic Interactions: Dynamic Constraints on Disordered TrkA Juxtamembrane Domain", *Journal of Physical Chemistry B*, 2019, 123, 50, 10709-10717.
23. Payel Mondal, Vishnu V. Krishnamurthy, Savanna R. Sharum, Neeka Haack, Huiwen Zhou, Jenifer Cheng, Jing Yang, **Kai Zhang*** "Repurposing protein degradation for optogenetic modulation of protein activities" *ACS Synthetic Biology*, 2019, 8, 11, 2585-2592 (Cover).
Highlighted in [Science Daily](#), [Nanowerk](#), [Illinois News Bureau](#)
24. I. Srivastava, J. S. Khamo, S. Pandit, P. Fathi, X. Huang, A. Cao, R. T. Haasch, S. Nie, **K.Zhang***, D. Pan* "Influence of Electron Acceptor and Electron Donor on the Photophysical Properties of Carbon Dots: A Comparative Investigation at the Bulk-State and Single-Particle Level. *Advanced Functional Materials* 2019, 1902466.
25. H. Hwang, Z. Jin, V. V. Krishnamurthy, A. Saha, P. S. Klein, B. Garcia, W. Mei, M. L. King, **K. Zhang**, J. Yang "Novel functions of the ubiquitin-independent proteasome system in regulating *Xenopus* germline development", *Development*, 2019, 146, 8, dev172700.
26. P. Fathi, J. S. Khamo, X. Huang, I. Srivastava, M. B. Esch, **K. Zhang***, D. Pan* "Bulk-state and single-particle imaging are central to understanding carbon dot photo-physics and elucidating the effects of precursor composition and reaction temperature", *Carbon*, 2019, 145, 572-585.

27. Q. Chen, X. Shao, Z. Tian, Y. Chen, P. Mondal, F. Liu, F. Wang, P. Ling*, W. He*, **K. Zhang***, Z. Guo, and J. Diao* “Nanoscale monitoring of mitochondria and lysosome interactions for drug screening and discovery”, *Nano Research*, 2019, 12, 5, 1009–1015.
28. B. Cai, L. Yu, S. R. Sharum, **K. Zhang***, J. Diao* “Single-vesicle measurement of protein-induced membrane tethering”, *Colloids and Surfaces B: Biointerfaces*, 2019, 177, 267-273.
29. J. S. Khamo, V. V. Krishnamurthy, Q. Chen, J. Diao, **K. Zhang***, “Optogenetic delineation of receptor tyrosine kinase subcircuits in PC12 cell differentiation”, *Cell Chemical Biology*, 2019, 26, 400-410.
Highlighted in [School of MCB](#) and [Neuroscience Program News](#) at UIUC.
30. S. K. Misra, I. Srivastava, J.S. Khamo, V. V. Krishnamurthy, D. Sar, A. S. Schwartz-Duval, J. A. N. T. Soares, **K. Zhang*** and D. Pan* “Carbon Dots with Induced Surface Oxidation Permits Imaging at Single-Particle Level for Intracellular Studies”, *Nanoscale*, 2018, 10, 18510-18519.
Highlight in the [School of MCB](#) at UIUC.
31. V. V. Krishnamurthy, **K. Zhang*** “Chemical physics in living cells – using light to visualize and control intracellular signal transduction” *Chinese Journal of Chemical Physics*, 2018 31(4), 375-392.
32. K. Sung, L. F. Ferrari, W. Yang, C. Chung, X. Zhao, Y. Gu, S. Lin, **K. Zhang**, B. Cui, M. L. Pearn, M. T. Maloney, W. C. Mobley, J. D. Levine and C. Wu ” Swedish Nerve Growth Factor Mutation (NGFR100W) Defines a role for TrkA and p75NTR in Nociception”, *Journal of Neuroscience*, 2018, 38(14), 3394-3413.
33. J.S. Khamo, V. V. Krishnamurthy, P. Mondal, S. R. Sharum, and **K. Zhang*** “Applications of optobiology in intact cells and multi-cellular organisms”, *Journal of Molecular Biology*, 2017, 429, 2999-3017.
34. V. V. Krishnamurthy, A. J. Turgeon, J. S. Khamo, W. Mei, P. Mondal, S. R. Sharum, J. Yang*, and **K. Zhang*** “Light-mediated, reversible modulation of protein localization and kinase activity during cell differentiation and *Xenopus* embryonic development” *Journal of Visualized Experiments (JoVE)*, 2017, 124, e55823.
35. Y. Osakada, **K. Zhang** “Single particle tracking reveals a dynamic role of actin filaments in assisting long-range axonal transport in neurons” *Bulletin of the Chemical Society of Japan (BCSJ)*, 2017, 90, 714-719.
36. P. Mondal, J. S. Khamo, V. V. Krishnamurthy, Q. Cai, and **K. Zhang*** “Drive the car(go)s— new modalities to control cargo trafficking in live cells” *Frontiers in Molecular Neurosciences*, 2017, 10, 4. doi: 10.3389/fnmol.2017.00004.
37. V. V. Krishnamurthy, J.S. Khamo, W. Mei, A. J. Turgeon, H. M. Ashraf, P. Mondal, D. B. Patel, N. Risner, E. E. Cho, J. Yang*, and **K. Zhang*** “Reversible optogenetic control of kinase activity during differentiation and embryonic development” *Development*, 2016, 143, 4085-4094.
38. V. V. Krishnamurthy, J. S. Khamo, E. Cho, C. Schornak, and **K. Zhang*** “Multiplex gene removal by two-step polymerase chain reactions”, *Analytical Biochemistry*, 2015, 481, 7-9.

39. V. V. Krishnamurthy, J. S. Khamo, E. Cho, C. Schornak, and **K. Zhang*** “Polymerase chain reaction-based gene removal from plasmids”, *Data in Brief*, 2015, 4, 75-82.

Contributed journal perspective and book chapter – independent career (UIUC)

40. **Kai Zhang***, Vishnu V. Krishnamurthy, “Enabling versatile control of molecular activity with small molecules and light”, *Journal of Molecular Biology*, 2020, 432, 19, 5209-5211.
41. Vishnu Krishnamurthy, John Khamo, Payel Mondal, Savanna Sharum, Jing Yang, and **Kai Zhang*** "Reversible Optogenetic Control of Growth Factor Signaling During Cell Differentiation and Vertebrate Embryonic Development", *Biophotonics Congress: Optics in the Life Sciences Congress*, 2019, AW1E.1.
42. V. V. Krishnamurthy, **K. Zhang*** “Simultaneous removal of multiple DNA segments by polymerase chain reactions” *Methods Mol Biol.*, Synthetic DNA, Ed R. Hughes. (Springer New York) 2017, 1472, 193-203.

Peer-reviewed journal publications – prior to UIUC

43. **K. Zhang*** and B. Cui* “Optogenetic control of intracellular signaling pathways”, *Trends in Biotechnology*, 2015, 33, 92-100. (*corresponding author)
44. P.D. Chowdary, D. Che, **K. Zhang**, B. Cui “Retrograde NGF axonal transport – coordination of opposite polarity motors near unidirectional motility regime” *Biophysical Journal*, 2015, 108, 2691-2703.
45. D. L. Che, L. Duan, **K. Zhang**, B. Cui, The dual characteristics of light-induced cryptochrome 2 homo-oligomerization and hetero-dimerization for optogenetic manipulation in mammalian cells, *ACS Synthetic Biology*, 2015, accepted.
46. L. Duan, D. Che, **K. Zhang**, Q. Ong, S. Guo, and B. Cui, Optogenetic control of molecular motors and organelle distributions in cells, *Chemistry & Biology*, 2015, 22, 671-682.
47. Q. Ong, S. Guo, L. Duan, **K. Zhang**, E. A. Collier, and B. Cui “The Timing of Raf/ERK and AKT Activation in Protecting PC12 Cells against Oxidative Stress”, *PLOS ONE*, 2016, e0153487.
48. Q. Ong, S. Guo, **K. Zhang**, and B. Cui “U0126 Protects Cells against Oxidative Stress Independent of Its Function as a MEK Inhibitor”, *ACS Chem. Neurosci.*, 2015, 6,130–137.
49. **K. Zhang** and B. Cui “Lighting up FGFR signaling”, *Chemistry & Biology*, 2014, 21, 806-808.
50. **K. Zhang**, L. Duan, Q. Ong, Z. Lin, P. Varman, K. Sung, and B. Cui “Light-mediated kinetic control reveals the temporal effect of the Raf/Mek/ERK pathway in PC12 cell neurite outgrowth”, *PLOS ONE*, 2014, 9, e92917.
51. **K. Zhang**, R. F. B. Kenan, Y. Osakada, W. Xu, R. S. Sinit, L. Chen, X. Zhao, J-Y. Chen, B. Cui, and C. Wu “Defective Axonal Transport of Rab7 GTPase Results in Dysregulated Trophic Signaling”, *J. Neuroscience* 2013, 33, 7451-7462.
52. W. J. Xie, **K. Zhang**, B. Cui “Functional characterization and axonal transport of quantum dot labeled BDNF”, *Integrative Biology*, 2012, 4, 953-960.

53. **K. Zhang**, Y. Osakada, W. J. Xie, and B. Cui “Automated image analysis for tracking cargo transport in axons”, *Microscopy Research and Technique* 2011, 74, 605-613.
54. K. A. Vossel, **K. Zhang**, X. Wang, G. Q. Yu, K. Ho, B. Cui, and L. Mucke “Tau reduction ameliorates $A\beta$ -induced impairments in axonal transport”, *Science* 2010, 330 198.
55. **K. Zhang**, H. V. Mudrakola, L. Chen, M. Vrljic, and B. Cui “Single molecule imaging of NGF axonal transport in a microfluidic device”, *Lab on a Chip* 2010, 10, 2566-2573.
56. H. V. Mudrakola*, **K. Zhang***, and B. Cui “Optically resolving individual microtubules in live axons using dynamic object tracking”, *Structure* 2009, 17, 1433-1441.
57. **K. Zhang**, W. K. Zhang, C. Y. Yang, and H. Yang “Bipolar Cellular Morphology of Malignant Melanoma in Unstained Human Melanoma Skin Tissue”, *J. Biomedical Optics* 2009, 14, 024042.
58. S. Li, **K. Zhang**, J. M. Yang, L. W. Lin, and H. Yang “Single Quantum Dots as Local Temperature Markers”, *Nano Lett.* 2007, 7, 3102-3105.
59. N. Ji, **K. Zhang**, H. Yang, and Y. R. Shen “Three-Dimensional Chiral Imaging by Sum Frequency Generation”, *J. Am. Chem. Soc.* 2006, 128, 3482-3483.
60. **K. Zhang**, H. Chang, A. H. Fu, A. P. Alivisatos, and H. Yang “Continuous Distribution of Emission States from Single CdSe/ZnS quantum dots”, *Nano Lett.* 2006, 6, 843-847.
61. **K. Zhang** and H. Yang “Photon-by-Photon Determination of Emission Burst from Diffusion Single Chromophores”, *J. Phys. Chem. B.* 2005, 109, 21930-21937.
62. **K. Zhang**, Z. J. Liu, and K. Y. Wang “Formation and Applications of Laser-Excited Surface Plasma Waves”, *Chinese J. Nature* 2002, 24, 44-47.

Contributed book chapter – prior to UIUC

63. **K. Zhang**, P.D. Chowdary, and B. Cui “Visualizing directional Rab7 and TrkA cotrafficking in axons by pTIRF microscopy” *Methods Mol Biol.*, 2015, 1298:319-29.
64. H. V. Mudrakola, C. Wu, **K. Zhang**, and B. Cui, “Single Molecule Imaging of Axonal Transport in Live Neurons”, in Laser Science XXV, OSA Technical Digest (CD) (Optical Society of America, 2009), LSThB3.
65. S. Li, **K. Zhang**, J-M Yang, L.W. Lin, and H. Yang “MEMS Temperature Characterization by CdSe Quantum Dots”, *The 14th International Conference on Solid-State Sensors, Actuators and Microsystems*, 2007, 1369-1372.
66. **K. Zhang**, N. Ji, Y. R. Shen, and H. Yang “Optically Active Sum Frequency Generation Microscopy for Cellular Imaging”, *Ultrafast Phenomena XV* Eds. P. Corkum, D. Jonas, D. Miller, A. M. Weiner, (Springer-Verlag, Berlin Heidelberg, 2007) 825.

INVITED TALKS AND PLATFORM IN CONFERENCE

1. Oh, T., Krishnamurthy V., Han, T., Zhang, K. “Elucidation of necroptosis pathway using Optogenetics”, 4th International Conference on Cell and Experimental Biology, Houston, Texas, April 2023

2. Mehta K., Yentch H., Lee J., Gao T.T., Zhang K, 'Phosphatidylinositol 3-phosphate mediates Arc capsids secretion through the multivesicular body pathway, Biophysical meeting, San Diego, February 2023
3. "Fine tune receptor tyrosine kinase activity by light", UWM Optical Microspectroscopy Symposium, Milwaukee, Wisconsin, August 2022.
4. "Single-Molecule Measurement of Orientation Distributions in Lipid Nanodiscs", UWM Optical Microspectroscopy Symposium, Milwaukee, Wisconsin, August 2022.
5. "Light up signal transduction – control cell fate with opsin-free optogenetics in health and disease", Chinese Biophysics Congress June 2022 (Virtual)
6. "Optogenetic control of body axis patterning during embryonic development", 2nd Optogenetic Technologies and Applications Conference, December 2021 (Virtual).
7. "Optogenetic control of neural differentiation and repair", Photopharmacology III, November 2021 (Virtual).
8. "Optogenetic control of neural differentiation, repair, and embryonic development", Chinese Bioscience Association, California, October 2021 (Virtual).
9. "Optogenetic activation of ERK and AKT signaling promotes axon regeneration in *Drosophila*", Biophysical Society 65th annual meeting, February 2021 (Virtual).
10. "Repurposing Protein Degradation for Optogenetic Modulation of Protein Activities" Society of Photo-Optical Instrumentation Engineers, San-Francisco, California, February 2020.
11. "Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development", Optogenetic Technologies and Applications, Boston, Massachusetts, December 2019. (**Session Chair**)
12. "Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development", Xenopus Resources and Emerging Technologies Meeting, Woods Hole, Massachusetts, October 2019.
13. "Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development", International Conference on Biomechanics and Medical Engineering (ICBME), San Diego, California, September 2019.
14. "Reversible optogenetic control of growth factor signaling during cell differentiation and Vertebrate Embryonic Development", OSA Biophotonics Congress, Tucson, Arizona, April 2019.
15. "Optogenetic activation and inactivation of the neurotrophin pathway in live cells" Experimental Biology, Orlando, Florida, April 2019.
16. "Temporal control of growth factor-mediated signaling pathways during cell differentiation and *Xenopus* embryonic development", American Society for Biochemistry and Molecular Biology Society Meeting, San Diego, California, April, 2018. (**Travel Award**)
17. "Using light to control the timing of kinase activity during cell differentiation and *Xenopus* embryonic development" Xenopus Resource and Emerging Technologies Meeting, Woods Hole Institute, Marine Biology Lab, Massachusetts, August 2017.

18. “Reversible optogenetic activation of neurotrophin-mediated signal transduction”, Houston Methodist Research Institute, Houston, Texas, March 2017.
19. “Reversible modulation of kinase activity during embryonic development”, Midwest Society of Developmental Biology Regional Meeting, Ann Arbor, Michigan, October 2016.
20. “Steering growth factor-mediated signal transduction by light”, East Lake Young Scholar Symposium, Huazhong University of Science and Technology, Wuhan, China, December 2014.
21. “Defective Axonal Transport of Rab7 GTPase Results in Dysregulated Trophic Signaling”, Bay Area Trafficking Symposium, UC Berkeley, California, September 2013.
22. “Impact of Charcot-Marie-Tooth type 2B disease-associated Rab7 mutations on signaling and axonal trafficking of NGF/TrkA”, 56th Biophysical Society Annual Meeting, San Diego, California, February 2012. [[Link](#)]

INVITED TALKS IN UNIVERSITY AND RESEARCH INSTITUTION

23. [Student Invited Talk] “Light It Up-Control Neuronal Functions via Opsin-free Optogenetics”, Undergraduate Neuroscience Society, UIUC, October 2023.
24. “Optical modulation of molecular activity during neural differentiation, regeneration, and embryonic development”, Department of biology, Penn State University, February 2021.
25. “Single-molecule detection, Super-resolution imaging, and optogenetics”, Biological Physics for Engineers invited lecture, University of Florida, February 2021.
26. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Chemistry, University of Alberta, September 2020.
27. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Physiology, Michigan State University, September 2020.
28. “Optical modulation of molecular activity during neural differentiation, regeneration, and embryonic development”, Department of Biochemistry, University of Illinois at Urbana-Champaign, IL, September 2020.
29. “Optical inhibition of growth factor signaling in living cells”, Cancer Center at Illinois, University of Illinois at Urbana-Champaign, IL, April 2020.
30. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Chemistry, University of Chicago, IL, March 2020.
31. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Chemistry, Princeton University, NJ, December 2019.
32. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Chemistry, University of Pennsylvania, PA, November 2019.

33. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Chemistry, Stanford University, CA, November 2019.
34. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Cancer Biology, University of Cincinnati, OH, October 2019.
35. “Bidirectional optogenetic control of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, Department of Bioengineering, University of California, San Diego, September 2019.
36. “Bidirectional optogenetic control of growth factor signaling during cell differentiation and embryonic development” Center for Physics of Living Cells, University of Illinois at Urbana-Champaign, IL, September 2019.
37. “Delineating receptor tyrosine kinase signaling pathways during cell differentiation and embryonic development” University of Southern California, Los Angeles, CA, December 2018.
38. “Developing an optogenetic toolbox for cell signaling control in mammalian cells and multicellular organisms” Center for Physics of Living Cells (CPLC), University of Illinois at Urbana-Champaign, Urbana, Illinois, July 2017.
39. “Dissection of growth factor signal transduction during cell differentiation and *Xenopus* embryonic development” Harvard Medical School, Boston, Massachusetts, June 2017.
40. “Dissection of growth factor signal transduction during cell differentiation and *Xenopus* embryonic development” Brown University, Providence, Rhode Island, June 2017.
41. “Delineating growth factor-regulated signaling pathways by light”, Department of Pathobiology of the College of Veterinary Medicine, University of Illinois at Urbana-Champaign, March 2017.
42. “Control the timing of the mitogen-activated protein kinase pathway during cell differentiation and *Xenopus* embryonic development”, School of Molecular Sciences, Arizona State University, Tempe, March 2017.
43. “Light-controlled growth factor signal transduction during cell differentiation and *Xenopus* embryonic development”, Department of Physiological and Molecular Plant Biology, University of Illinois at Urbana-Champaign, Urbana, Illinois, January 2017.
44. “Developing an optogenetic toolbox for cell signaling control” Center for Physics of Living Cells, University of Illinois at Urbana Champaign. July 2016.
45. “Study signal transduction in live cells by light”, School of Molecular and Cellular Biology, University of Illinois at Urbana-Champaign, Urbana, Illinois, August 2015.
46. “Control cell fate determination by light”, Center for Biophysics and Computational Biology, University of Illinois at Urbana-Champaign, Urbana, Illinois, August 2015.
47. “Control PC12 cell differentiation by light”, Neuroscience program, University of Illinois at Urbana-Champaign, Urbana, Illinois February 2015.

48. “Steering growth factor-mediated signal transduction by light”, Fudan University, Shanghai, China, December 2014.
49. “Steering growth factor-mediated signal transduction by light”, Huazhong University of Science and Technology, Wuhan, China, December 2014.
50. “Light-controlled activation of the mitogen-activated protein kinase pathway”, Center for Biophysics and Computational Biology, University of Illinois at Urbana-Champaign, Urbana, Illinois, July 2014
51. “Observation and modulation of signal transduction in live cells using light”, California Institute of Technology, California, January 2013.
52. “Dysregulated axonal transport of NGF/TrkA in Charcot-Marie-Tooth type 2B disease” Biophysics Talks, Stanford University, Stanford, California, January 2012.
53. “Observing quantum dot one at a time: optical characterization and applications in live cell imaging”, Peking University, P. R. China, November 2011.
54. “Single Chromophore Experiments and Quantitative Analysis”, November 23, 2006, Zhengzhou University, Zhengzhou, Henan, P. R. China.

POSTER PRESENTATIONS AT CONFERENCE

55. Kangqiang Qiu, Weiwei Zou, Hongbao Fang, Minggang Hao, Kritika Mehta, Zhiqi Tian, Jun-Lin Guan, Taosheng Huang, Jiajie Diao, **Kai Zhang**, “Optical modulation of mitochondrial morphology and functions” 67th Biophysical Society Meeting, San Diego, California, February 2023.
56. S. R. Sharum, P. Mondal, V. V. Krishnamurthy, K. Mehta, H. Fan, J. Yang, **K. Zhang** “Optical modulation of receptor tyrosine kinase signaling during cell differentiation and embryonic development”, 64th Biophysical Society Meeting, San Diego, California, February 2020.
57. Z. Wang, H. Fan, X. Hu, J. Khamo, J. Diao, **K. Zhang**, T. Pogorelov, “Coaction of electrostatic and hydrophobic interactions in signaling: dynamic constraints on disordered TrkA juxtamembrane domains”, 64th Biophysical Society Meeting, San Diego, California, February 2020.
58. P. Mondal, V. V. Krishnamurthy, S. R. Sharum, N. Haack, J. Yang, **K. Zhang** “Repurposing protein degradation for optogenetic modulation of protein activities”, SPIE Photonic West Bio, San Diego, California, February 2020.
59. N. Haack, P. Mondal, V. V. Krishnamurthy, S. R. Sharum, **K. Zhang** “Repurposing protein degradation for optogenetic modulation of protein activities”, Society for Neuroscience, Chicago, Illinois, December 2019.
60. V. V. Krishnamurthy, J. Khamo, P. Mondal, S. R. Sharum, **K. Zhang** “Bidirectional control of receptor tyrosine signaling during cell differentiation with light”, Society for Developmental biology annual meeting, Boston, Massachusetts, July 2019.
61. V. V. Krishnamurthy, J. Khamo, P. Mondal, S. R. Sharum, **K. Zhang** “Bidirectional optogenetic control of growth factor signaling during cell differentiation and vertebrate

- embryonic development”, Gordon Research Conference, Salve Regina University, Newport, Rhode Island, June 2019.
62. S. R. Sharum, P. Mondal, K. Cho, **K. Zhang** “Temporal inhibition of ERK Activity by Optogenetic Control of MAPK Phosphatase 3” Experimental biology, Orlando, Florida, April 2019.
 63. J. Khamo, **K. Zhang** “Optogenetic delineation of receptor tyrosine kinase subcircuits in pc12 cell differentiation”, 63th Biophysical Society Meeting, Baltimore, Maryland, March 2019.
 64. P. Mondal, V. V. Krishnamurthy, J. Khamo, J. Yang, **K. Zhang** “Temporal control of growth factor-mediated signaling pathways during cell differentiation and *Xenopus* embryonic development”, American Society for Biochemistry and Molecular Biology Society Meeting, San Diego, California, April 2018. (**Travel Award**)
 65. **K. Zhang** “Control neurotrophin signaling using light during PC12 cell differentiation and *Xenopus* embryonic development”, Biophysical Society Meeting, San Francisco, California, February 2018. [[Link](#)]
 66. **K. Zhang** “Dissection of growth factor-regulated signaling pathways by light”, Society for Developmental biology annual meeting, Minneapolis, Minnesota, July 2017.
 67. **K. Zhang** “Control neurotrophin signaling using light during cell differentiation and *Xenopus* embryonic development”, Gordon Research Conference, Salve Regina University, Newport, Rhode Island, June 2017.
 68. **K. Zhang** “Developing an optogenetic toolbox for cell signaling control”, Center for Physics of Living Cells, University of Illinois at Urbana-Champaign, Urbana, Illinois, July 2016
 69. **K. Zhang** “Resolving intracellular mechanisms of neurotrophin-mediated signal transduction via optogenetics” 16th International symposium on neural regeneration (ISNR), Pacific Grove, California, December 2015.
 70. Q. Ong, A. McGuire, S. Guo, F. Santoro, **K. Zhang**, and B. Cui “Optogenetic spatial control of TrkA-mediated pathways reveals a potential role for Raf/ERK pathway in inducing polarity in PC12 cell differentiation model” American Society for Cell Biology ASCB, San Diego, California, December 2015.
 71. **K. Zhang** “Light-controlled growth factor-mediated signal transduction”, 59th Biophysical Society Annual Meeting, Baltimore, Maryland, February 2015.
 72. Q. Ong, **K. Zhang**, S. Guo, L. Duan, and B. Cui “Optogenetic modulation of the Raf/ERK pathway in PC12 cells”, ASCB local meeting, Quantitative Imaging in Cell Biology, Santa Clara University, California, May 2014 (**Best poster award**).
 73. **K. Zhang**, L. Duan, Q. Ong, Z. Lin, P. Varman, K. Sung, and B. Cui “Light-controlled MAPK signaling pathway reveals a memory effect in PC12 cell neurite outgrowth”, Single Cell Analysis Symposium, Stanford University, California, September 2013.
 74. **K. Zhang**, L. Duan, Z. Lin, K. Sung, Y. Osakada, and B. Cui “Control the mitogen-activated protein kinase signaling pathway by light”, Synthetic Biology Gordon Research Conference, Mount Snow Resort, Vermont, June 2013.

75. **K. Zhang**, L. Duan, Z. Lin, K. Sung, Y. Osakada, and B. Cui “Light-controlled mitogen - activated protein kinase (MAPK) signaling pathway in live cells”, 57th Biophysical Society Annual Meeting, Philadelphia, Philadelphia, February 2013.
76. W. Zhao, **K. Zhang**, W. Xie, L. Hanson, Z. Lin, Y. Cui, and B. Cui “Accelerating the development of hippocampal neurons using nanopillar structures”, 57th Biophysical Society Annual Meeting, Philadelphia, February 2013.
77. **K. Zhang**, L. Duan, Z. Lin, K. Sung, Y. Osakada, and B. Cui “Precise control of signal transduction in living cells by light”, 2012 American Society for Cell Biology Annual Meeting, San Francisco, California, December, 2012.
78. **K. Zhang**, Y. Osakada, M. Vrljic, L. Chen, H. Mudrakola, and B. Cui “Single-molecule imaging of nerve growth factor axonal transport in microfluidic devices”, 55th Biophysical Society Annual Meeting, Baltimore, Maryland, March 2011.
79. **K. Zhang**, C. Wu, H. Mudrakola, Y. Osakada, and B. Cui “Real time visualization of axonal transport of GTPase Rab7 in rat embryonic dorsal root ganglia”, 54th Biophysical Society Annual Meeting, San Francisco, California, February 2010.
80. Y. Osakada, H. Mudrakola, **K. Zhang** and B. Cui “Effects of actin filaments on NGF retrograde transport”, 54th Biophysical Society Annual Meeting, San Francisco, California, February 2010.
81. **K. Zhang**, W. K. Zhang, C. Y. Yang, and H. Yang “Nonlinear optical imaging of melanocytes in collagen matrix”, 234th American Chemical Society National Meeting & Exposition, Boston, Massachusetts, August 2007.
82. H. Yang, **K. Zhang**, A. Fu, P. Alivisatos, C. Hayden “Studying Photoluminescence Dynamics of Single Quantum Dots Photon by Photon”, American Physical Society Meeting, Baltimore, Maryland, March 2006.
83. **K. Zhang** and H. Yang “Photon-by-photon determination of emission bursts from diffusing single chromophores”, American Physical Society Meeting, Baltimore, Maryland, March 2006.
84. **K. Zhang** and H. Yang “Photon-by-photon determination of emission bursts from diffusing single chromophores”, 231st American Chemical Society Meeting & Exposition, Atlanta, Georgia, March 2006.
85. N. Ji, **K. Zhang**, H. Yang, and Y. R. Shen “Sum frequency generation microscopy for imaging chirality”, 50th Biophysical Society Annual Meeting, Salt Lake City, Utah, February 2006.
86. **K. Zhang** and H. Yang “Field and fluorescence modification by colloidal gold nanoparticles”, Materials Research Society Spring Meeting, San Francisco, California, March 2005.
87. **K. Zhang**, H. Chang, A. H. Fu, L. P. Watkins, A. P. Alivisatos, and H. Yang “Photon by photon analysis of single quantum dot emission dynamics”, Materials Research Society Spring Meeting, San Francisco, California, March 2005.

TEACHING EXPERIENCE**Instructor****University of Illinois at Urbana-Champaign**

Physical Biochemistry (MCB/BIOC 446, CHEM 472)	2016-present
Neuroscience Program (NEUR598, Organizer: Justin Rhodes)	2016-present
Grading Faculty for Scientific Writing (MCB 540)	2023 Spring
Center for Physics of Living Cells, Summer Workshop	2016-present
Tutorial (BIOP 586)	2015-present

Students: Siqu Liao (Lu), Congjian Ni (Lu) Yongjae Lee (Selvin), Chang-Ting Lin (Ha), Kai Wen Teng (Selvin), Zhiyu Zhao (Tajkhorshid), Chaoyi Jin (Selvin), Brooke Ramsey (Sangjin Kim)

Part-time Lecturer**Biomedical, Chemical and Materials Engineering Department****San Jose State University**

Graduate-division Chemical Engineering Thermodynamics	2012
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Teaching Assistant**Department of Chemistry, UC Berkeley**

Graduate-division Chemical Kinetics	2005
Undergraduate General Chemistry	2004
Undergraduate General Chemistry	2003

PROFESSIONAL ACTIVITIES**Ad Hoc journal reviewer****Biological Sciences:**

Science, Nature Cell Biology, Nature Communications, Nature Protocols, Trends in Genetics, Cell Chemical Biology, Journal of American Chemical Society, ACS Synthetic Biology, Development, Journal of Molecular Biology, ACS Chemical Neuroscience, ACS Chemical biology, Expert Opinion on Drug Discovery, Journal of Integrative Neurosciences, Scientific Reports, Methods, Frontiers in Molecular Neuroscience, Frontiers in Cell and Developmental Biology, ChemBioChem, Journal of Visualized Experiments, Dyes and Pigments, Experimental Neurology

Physical Sciences:

Journal of Physical Chemistry B, Journal of Physical Chemistry Letter, Chemical Sciences, Journal of Biomedical Optics

Engineering and Materials Sciences:

Nature Biomedical Engineering, Nature Methods, Advanced Healthcare Material, Current Opinion in Biotechnology, Advanced Materials, Journal of Micro/nanolithography, MEMS, and MOEMS (J3M), SLAS Technology, Small

Mentoring

Postdoctoral Scholar

Dr. Vishnu Vardhan Krishnamurthy 2019 – 2020
 Current position: Director, High-throughput Screening Facility
 Dr. Tyler Camp (American Parkinson Association Postdoctoral Fellow) 2021 - present

Ph.D. students

Dr. John Khamo 2014-2019
 Robert L. Switzer Outstanding TA (2019), Keith Westcott Fellow (2017)
 Current position: Scientist at Situ Biosciences (2021) and Neelyx lab (2022).

Dr. Vishnu Vardhan Krishnamurthy 2014 - 2019
 L. Switzer Outstanding TA (2018) Leading author of a Featured Communication and Front Cover in the Journal of Molecular Biology (2021).
 Current position: Director at UIUC HTSF

Dr. Payel Mondal 2015 - 2020
 ASBMB Travel Award (2018), Dissertation Completion Fellowship (2019), Switzer Outstanding TA (2020). Anne A. Johnson Work Award (2021, delayed from 2020).
 Current position: Postdoc at Prof. CHRISTINA TOWERS' laboratory, Salk Institute

Dr. Savanna Sharum Skeeters 2016 - 2022
 Keith Westcott Fellow (2018), second prize in the 1st School of MCB retreat (2019), Robert L. Switzer Outstanding TA (2019).
 Current position: Scientist at Cyrus Biotech

Graduate Students

Huaxun Fan (Kade Fellow, 2021)
 Kritika Mehta (Kade Fellow, 2022, Robert L. Switzer Outstanding TA 2021,2022)
 Teak-Jung Oh (Korean American Scholar Fellow, LAS Travel Award 2023)
 Andrew Huang (NSF GRFP Honorable Mention, Biophysics Graduate Fellow, 2022)
 Bryan Gworek (Biophysics, 2023)
 Amna Mehfooz (Biochemistry, 2023)
 Zixiao Li (Biochemistry, 2023)

Supervisor of undergraduate students in the laboratory and current position

Cara Schornak	Graduate student in Vanderbilt University.	2014-2015
Ellen Cho	Biochemistry high distinction, work at Loyola University Chicago	2014-2016
Humza Ashraf	Biochemistry high distinction, Summer Research Award, Graduate student at the University of Colorado, Boulder.	2015-2017
Dil Patel		2015-2017
Noah Risner		2015-2017
Adam Barm		2015-2015
Neeke Haack		2015-2019
Jennifer Cheng,	Biochemistry highest distinction, Outstanding Student, Outstanding Thesis Award, Graduate student in Northwestern University	2017-2018
Kelly Cho	MCB Summer Undergraduate Research Fellowship, Medical School student at University of Illinois Chicago	2017-2019
Rachel Benedeck		2018-2018
Huiwen Zhou	Graduate student at Northwestern University	2018-2019
Max Spiro		2019-2020
Dhananjay Shahdadpuri		2019-2020
Tina Han (SURF fellowship)		2020-2022
Irina Cheng (Northwestern University Summer Undergraduate Research Fellow)		2020-2021
Collin Barnes (BEST Fellow of MCB)		2021-present
Uma Agarwal, Highest Distinction, Thesis Award		2021-2023
Nicholas Worley		2021-2022
Aruniti Manawa		2021-2022
Ami Patel		2021-2023
Tristan Worley		2021-2023
Nathan Gron, Highest Distinction, Thesis Award		2021-2023
Nafay Abdul, SURF 2022,2023		2021-2023
Ahnaf Monyem		2021-2022
Kurt Leano, SURF 2023		2022-2023
Danyi Wang, SURF 2023		2022-2023
Yushan Li		2022-2023
Jillian Leturno		2023-2023
Jungbin Lee, (James Scholar, SURF 2022)		2022-present
Zayn Beg, SURF 2023		2022-present
Henry Yentsch		2022-present
Meghan McCann, SURF 2023, Sweden exchange program		2023-present
Asher Bhurgri		2023-present

Advisor for first-year rotation graduate student in MCB

2014	John Khamo, Erik Andersen
2015	Payel Mondal, Eric Shinn, Yeoan Youn
2016	Madhura Duttagupta, Nandan Haloi, Savanna Sharum
2018	Huaxun Fan, Micca Hecht, Kritika Mehta, Laura Sutenfield
2020	Jessica Palalay, Anna Mankovich, Kaylee Kuzelka, Jorge Calderin
2021	Ran Yang, Anuradha Sharma, Andrew Huang (Biophysics), AnuTanner, Terry Gao, Xiangning Song, Dean (Biophysics) Tanner, Rebecca Tang, Shweta Shree

2022 Sepehr Alaeen (Biophysics), Suhail Chhakara, Katrine Dailey
 2023 Zixiao Li, Bryan Gworek (Biophysics), Gabe McKenna, Harrison Wu (Biophysics), Adriana Andrus, Julius Benson (Biophysics), Amna Mehfooz, Rujuta Pendharkar (Biophysics)

Faculty advisor of other Biochemistry undergraduate students

Junyao Zhu, Zhouyang Zhu, Haoyuan Yan, Luis Tadeo, Tyanporn Tangpradabdul 2017-2018

Supervisor of high school students in the laboratory and current position

Ana Rosu Johns Hopkins University 2018-2020
 Sana Nabi Herricks High School (New York) Virtual 2021

Outside Ph.D. dissertation committee

Arizona State University (Faculty: Prof. Jia Guo) 2017
 University of Alberta (Student: Ms. Xiaocen Lu, Faculty: Prof. Robert E. Campbell) 2020
 Nanyang Technological University, Singapore
 (Student: Huanwen Mu, Xinwen Miao; Faculty: Prof. Wenting Zhao) 2022-2023

Memberships

Society of Developmental Biology 2016-present
 Biophysical Society 2005-present
 American Society for Cell Biology 2012
 Optical Society of America 2009
 American Chemical Society 2003 - 2006
 Material Research Society 2003 - 2006
 American Physical Society 2003 - 2006

Service

Strategic Advisory Committee, MCB, UIUC 2022-present
 Biophysics Executive Committee, UIUC 2022-present
 Biophysics Entrance Advisory Committee, UIUC 2023-present
 Neuroscience Program Award Committee, Chair & Executive Committee, UIUC 2023-present
 LAS General Education Committee, UIUC 2022-2023
 Admission Committee, Biophysics, UIUC 2018-present
 MCB Retreat Committee, UIUC 2023
 MCB Admission committee, UIUC 2023
 General Education Committee, LAS, UIUC 2023
 Faculty Search Committee, MCB, UIUC 2020, 2023
 Course and Curriculum Committee, MCB, UIUC 2020 - 2021
 Seminar Committee, Biochemistry, UIUC 2016 - 2021
 BEST Scholarship Committee, MCB, UIUC 2020

Outreach

Instructor

Center for Physics of Living Cells (NSF funded) Summer workshop, <i>UIUC</i>	2016 - 2020
Next generation Science Technology Engineering Art Math (STEAM) demonstration (Nano Class 3-5 grade)	studio science 2017
Ecole Bilingue de Berkeley primary school Second grade, Berkeley, California	2017
Judge MCB Undergraduate Research Symposium, <i>UIUC</i>	2021
Undergraduate research conference	
East Central Illinois American Chemical Society, <i>UIUC</i>	2016
Synopsys Championship	
<i>Santa Clara Valley Science and Engineering Fair Association</i>	2009
Co-founder Undergraduate Student Travel Award	
<i>University of Science and Technology of China</i>	2008 - 2011