



J. Plankton Res. (2014) 36(2): 326–333. First published online January 20, 2014 doi:10.1093/plankt/fbt136

HORIZONS

Disease in freshwater zooplankton: what have we learned and where are we going?

CARLA E. CÁCERES^{1,2*}, ALAN J. TESSIER³, MEGHAN A. DUFFY⁴ AND SPENCER R. HALL⁵

¹PROGRAM IN ECOLOGY, EVOLUTION AND CONSERVATION BIOLOGY, UNIVERSITY OF ILLINOIS AT URBANA-CHAMPAIGN, URBANA, IL 61801 USA, ²SCHOOL OF INTEGRATIVE BIOLOGY, UNIVERSITY OF ILLINOIS AT URBANA-CHAMPAIGN, URBANA, IL 61801, USA, ³NATIONAL SCIENCE FOUNDATION, DIVISION OF ENVIRONMENTAL BIOLOGY, 4201 WILSON BLVD, ARLINGTON, VA 22230, USA, ⁴DEPARTMENT OF ECOLOGY AND EVOLUTIONARY BIOLOGY, UNIVERSITY OF MICHIGAN, ANN ARBOR, MI 48109, USA AND ⁵DEPARTMENT OF BIOLOGY, INDIANA UNIVERSITY, BLOOMINGTON, IN 47405, USA

*CORRESPONDING AUTHOR: 515 Morrill Hall, Urbana, IL 61801. Tel: +217-244-2139. E-mail: caceres@life.illinois.edu

Received September 23, 2013; accepted December 21, 2013

Corresponding editor: Beatrix E. Beisner

In the last few decades, zooplankton (especially *Daphnia*) have emerged as a model system for examining the ecological and evolutionary roles of parasites in populations, communities and ecosystems. We build on this foundation, moving towards continued integration of epidemiology and community ecology to understand the distribution and abundance of infectious diseases. Future studies should link processes that occur over multiple time scales: from deep-time phylogenetic patterns that have shaped and been shaped by host–parasite interactions, to the dynamics that play out over decades or within seasons, to the physiological time scales over which parasites operate to infect and kill their hosts. We also consider how the combination of processes across multiple time scales has shaped the biogeography and changing mosaic of host–parasite interactions in time and space. Finally, we consider a future where zooplankton–parasite interactions are influenced by global change. The *Daphnia*–parasite model system offers a powerful tool to link anthropogenic changes to infectious diseases, including zoonotic diseases of significance to human health.

KEYWORDS: *Daphnia*; host; parasite

INTRODUCTION

Parasites and pathogens can severely depress their host populations, and their ecological impact seems destined to grow as global climate change, species invasions and anthropogenic disturbance intensify (Daszak *et al.*, 2000; Brearley *et al.*, 2012). Freshwater zooplankton (especially *Daphnia*) have increasingly been used to demonstrate connections between ecology, evolution and the distribution and abundance of infectious diseases. Our intent here is not to recreate some excellent recent reviews on these topics (Ebert, 2005, 2008; Lampert, 2011; Miner *et al.*, 2012). Rather we seek to highlight how broad insights gleaned from *Daphnia*–parasite interactions have advanced our understanding of the ecology and evolution of host–parasite interactions. We also point to the frontiers of research using zooplankton systems. We see great opportunities to use plankton to create highly integrative insight into disease. This insight has the potential to move across scales of biology, from within-host physiological and immunological interactions to population and food web dynamics to ecosystem processes (Fig. 1). Work with plankton disease can also

facilitate integration of physiological, ecological and evolutionary mechanisms operating across time scales.

WHAT MAKES A MICROBE A PATHOGEN FOR SOME BUT NOT ALL SPECIES OF ZOOPLANKTON?

The evolution of pathogen specificity in infection is a major unanswered question in the infectious disease literature. The dozen or so commonly documented parasites of freshwater zooplankton show a wide spectrum of specificity, ranging from those that infect only certain genotypes of certain species to those infecting a wide range of genotypes and species (Stirnadel and Ebert, 1997; Bittner *et al.*, 2002; Ebert, 2005; Wolinska *et al.*, 2006; Altermatt and Ebert, 2007; Ben-Ami *et al.*, 2008; Duffy *et al.*, 2010; Lampert, 2011). What are the mechanisms underlying this variability? The answer to this question surely depends on traits of both parasite and host. *Daphnia*, like all invertebrates, have an immune system (Mucklow and Ebert,

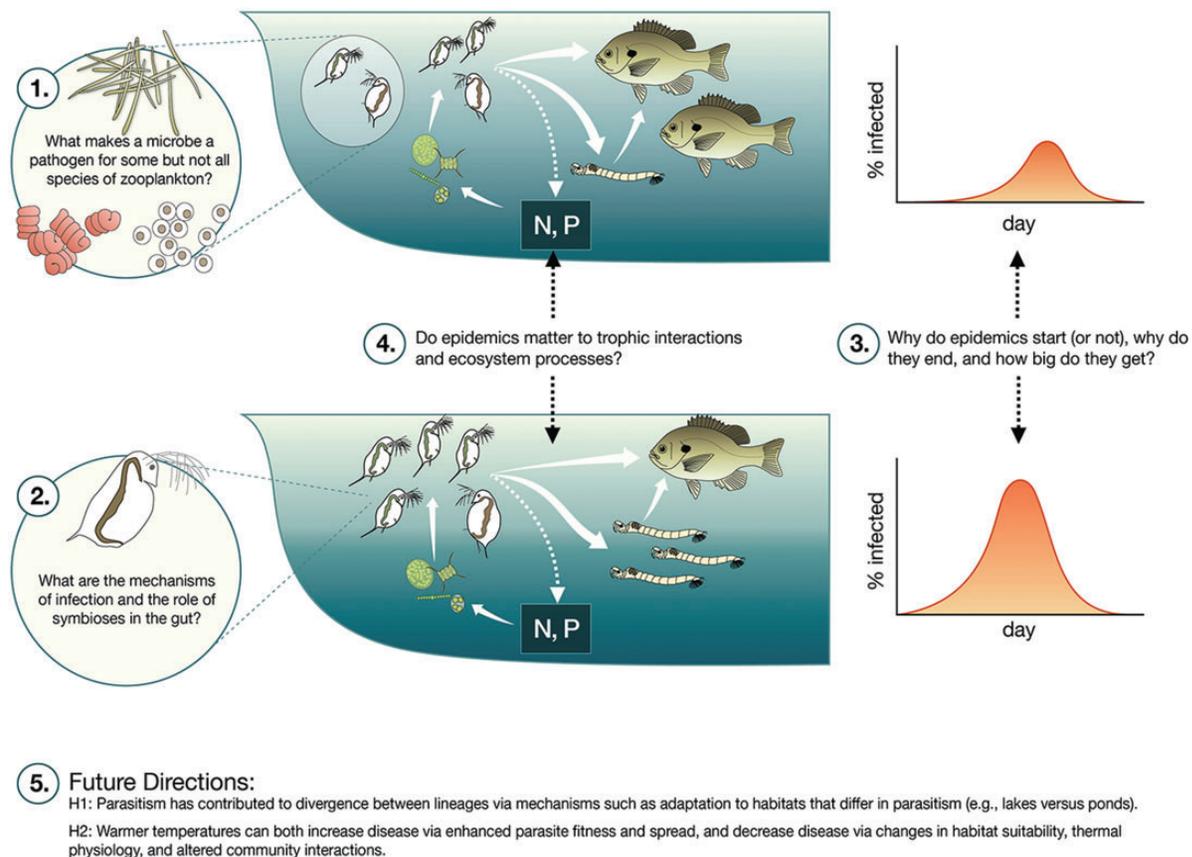


Fig. 1. A conceptual diagram of several of the major concepts described in the text. Numbers correspond to the five major sections of the paper. We list only two hypotheses as examples. The figure depicts two thermally stratified lakes that differ in key habitat features (e.g. strength of stratification) and the relative abundance of key food web players (diluters and predators) that are known to influence the timing and magnitude of epidemics (see also Hall *et al.*, 2010). Figure drawn by John Megahan.

2003; McTaggart *et al.*, 2009; Decaestecker *et al.*, 2011), which in some cases appears ineffective (Auld *et al.*, 2010). What determines whether a particular microbe can evade the immune system of a particular host? Little is known about the links between mechanisms of pathogenicity and host specialization. However, answers to this question are likely to help explain why, in some cases, there is clear evidence for ‘local adaptation’ (where parasites are most virulent on hosts from the same population; Ebert, 1994; Ebert and Hamilton, 1996; Refardt and Ebert, 2007), whereas, in others, there is no evidence for local adaptation (Duffy and Sivars-Becker, 2007; Searle *et al.* in prep.).

Future research tackling the drivers of host–parasite specificity should focus on the genetic and environmental factors involved in infection and the subsequent evolution of virulence. Flagship work on the *Daphnia magna*–*Pasteuria ramosa* system has provided some answers at the within-species (genotype) level (Ebert *et al.*, 2004; Mitchell *et al.*, 2005; Jensen *et al.*, 2006; Bieger and Ebert, 2009; Ben-Ami *et al.*, 2011; Hall and Ebert, 2012). Strong genetic specificity of hosts and parasites (Carius *et al.*, 2001; Luijckx *et al.*, 2012) provide an excellent model to uncover mechanisms of resistance to parasitism. However, such specificity is not universal (Duffy and Sivars-Becker, 2007; Auld *et al.*, 2012; Searle *et al.* in prep.). Hence, comparative studies on other zooplankton–parasite systems could provide more insight into within-host mechanisms that govern defense and virulence. It could be particularly interesting to use *Daphnia*–parasite systems that differ in the degree of specificity to address the genetic architecture of parasite susceptibility (see review by Wilfert and Schmid-Hempel, 2008 for how the genetic architecture of susceptibility has been addressed in other systems).

Complementary insight could be gleaned from study of the phylogenetic relationships of Daphniidae and their parasites. Divergent patterns of coevolution, revealed from a deeper time perspective of diversification, might shed light on variation in specificity. We know that evolution of life histories of daphniids depends on multiple selective factors (e.g. predation, resources, competitors) and habitat type (e.g. pond vs. lake) (reviewed in Lampert and Sommer, 2007; Lampert, 2011). Have these various selective factors influenced the evolution and distribution of parasites among daphniid species? Plankton offer great advantages to tackle this problem. Unlike in many other systems, plankton ecologists can readily exploit three key factors, cyclical parthenogenesis, tendency to hybridize and apparent pond–lake divergence into sister species, to identify and disentangle genetic and environmental mechanisms of defense within the context of recent evolutionary divergence by habitat.

However, a (temporary) problem impedes progress on this front. Although many daphniid species have broad

ranges, we know little about the biogeography of parasites within different host species. Comparisons within and among clades over broad geographic domains could produce exciting results. To illustrate, consider two important species of pond *Daphnia*. Why are parasites so rare among *D. pulex* in North America yet common in *D. magna* in Europe (Ebert, 2005; Killick *et al.*, 2008; Cáceres *et al.* unpublished data)? Both species live in shallow, often ephemeral water bodies that lack vertebrate predators. Both species have close contact with the sediment and potentially the parasite spores therein (Decaestecker *et al.*, 2002). Does some physiological or ecological adaptation to pond habitat constrain disease emergence in *D. pulex* but not in *D. magna*?

Finally, recent invasions of host species, coupled with ‘resurrection’ approaches (where it is possible to retrieve the dormant stages from both the host and the parasite over multiple years), can be exploited to study adaptation and possible co-evolution (Decaestecker *et al.*, 2007, 2013). The invasion of *Daphnia lumholtzi* into North America (Havel *et al.*, 2000) and its subsequent infection by *Pasteuria* (M.A. Duffy, unpubl. data) is one such example. Such studies, when coupled with an examination of deep-time evolutionary diversification can help to address questions related to the mechanisms underlying pathogen specificity and virulence.

WHAT ARE THE MECHANISMS OF INFECTION AND THE ROLE OF SYMBIOSES IN THE GUT?

The gut of zooplankton hosts lies at an intersection of resistance, microbiology and physiology. Many pathogens enter the zooplankton host’s body via the gut (Ebert, 2005). What defense mechanisms operate in the gut and do they tradeoff with resource acquisition? One type of defense may involve microbes, but we are just beginning to understand the *Daphnia* microbiome (Freese and Schink, 2011). The human microbiome project has indicated that our gut flora can influence disease dynamics within human hosts (Foxman and Goldberg, 2010), and symbionts of other invertebrates can confer resistance to disease (Parker *et al.*, 2013). Can the gut microbiome of *Daphnia* provide critical defense and influence variation in resistance to infection? Theory predicts that the host’s symbiont community may facilitate or hinder the evolution of pathogen specialization, yet this theory remains untested for most systems (Antonovics *et al.*, 2013). Furthermore, can changes in microbial gut community influence the seasonal timing of disease outbreaks? Are some pathogens only facultatively pathogenic? If so, which conditions determines their harm or benefit?

The gut may also provide the link between disease, gut physiology and foraging ecology. For instance, gut passage time varies greatly among daphniids (ranging from 5 to >25 min) and this variation reflects differences in body size, gut volume and feeding rate among species and life stages (DeMott *et al.*, 2010). It remains unknown if gut passage time influences susceptibility to pathogens acquired while feeding. Resources can fundamentally alter both host density and key epidemiological traits such as production of infectious propagules (spores) and susceptibility and tolerance to infection (Pulkkinen and Ebert, 2004; Hall *et al.*, 2009a, b, 2010, 2012; Frost *et al.*, 2008a; Civitello *et al.*, 2013). Increased research linking *Daphnia* foraging behavior, physiology and ecology to susceptibility to various species of pathogens is needed to resolve these questions. Such integrative research provides a means of linking mechanisms governing individual host–parasite interactions to population scale dynamics and food web interactions.

WHY DO EPIDEMICS START (OR NOT), WHY DO THEY END, AND HOW BIG DO THEY GET?

There are numerous case studies documenting a repeatable pattern of seasonal increases in prevalence of infection (Lass and Ebert, 2006; Cáceres *et al.*, 2006; Johnson *et al.*, 2009). But what determines when epidemics start, when they end, and how big they get? Epidemiological theory tells us that a parasite will invade its host population and spread when the basic reproductive ratio of the parasite (R_0) exceeds 1. But this invasion success hinges on a variety of factors including climate, physical mixing processes, host behavior, interspecific interactions and levels of host resistance (which may evolve rapidly).

Water movements may trigger the onset of disease in some zooplankton disease systems (e.g. the fungus *Metschnikowia bicuspidata* in *Daphnia dentifera*; Hall *et al.*, 2010). But important questions remain unanswered. Can too much mixing inhibit epidemics by reducing the encounter of host and parasite? Will global change influence disease by altering seasonal mixing patterns? Furthermore, not all diseases seem to require mixing for epidemics to begin. For instance, epidemics of a chytrid (*Polycaryum laeve*) reached peak prevalence in *Daphnia* under the ice (Johnson *et al.*, 2006a).

Habitat selection behavior (both in space and time) is another likely candidate for determining disease transmission. Host aggregation during vertical migration can influence transmission dynamics, as exemplified in the ‘Deep Trouble’ hypothesis (Decaestecker *et al.*, 2002). *Daphnia* that remain near sediments to avoid visual predators in ponds may increase their encounter rate with parasite spores in

sediments. Diapause, both seasonal and prolonged, provides another potentially interesting form of habitat selection by hosts (Hairston and Cáceres, 1996; Cáceres, 1997). Habitat selection theory has largely focused on predation and resource competition and has not fully considered the potential role of parasites (e.g. Leibold, 1990, 1991; Leibold and Tessier, 1991). More generally, how do parasites interact with predators and competitors to determine the spatial and temporal distribution of host and parasite genotypes and species within stratified lakes? How have parasites influenced, or been influenced by host adaptations to the benthic vs. pelagic habitat? One general goal is to understand the net effects of physical processes and host aggregation behavior (which can be influenced by mixing and stratification) in determining encounter and transmission rates.

Interspecific interactions play an important role in determining the outcome of host–parasite interactions. Predators can directly influence disease spread in freshwater zooplankton either by culling infected hosts or by spreading infected propagules while feeding (Duffy *et al.*, 2005; Johnson *et al.*, 2006b; Pulkkinen and Ebert, 2006; Cáceres *et al.*, 2009; Hall *et al.*, 2010). Moreover, both planktivorous fish and *Chaoborus* have well-known indirect effects on their *Daphnia* prey (Tollrian, 1993; Boersma *et al.*, 1998; Reissen, 1999) and these behavioral, morphological and life history changes have the potential to both increase and decrease transmission (Yin *et al.*, 2011; Duffy *et al.*, 2011; Bertram *et al.*, 2013). For a particular host–parasite system, do these direct and indirect effects of predation act in the same direction (thus enhancing the effect of a predator on disease dynamics) or act in opposite directions (making the predator effect more difficult to detect)? Competition may have equally important effects on the distribution and prevalence of disease. Multiple studies have also indicated that the presence of a parasite can reverse competitive rankings among hosts (Wolinska *et al.*, 2006; Bieger and Ebert, 2009; Refardt and Ebert, 2012). Does this interaction between competition and parasitism underlie the temporal habitat selection (seasonal succession) that is so commonly observed in freshwater zooplankton? For example, can a population’s seasonal phenology be driven (at least in part) by parasite—mediated selection for the timing to enter or terminate dormancy (Duncan *et al.*, 2006)?

Basin size and shape, which structures trophic interactions, may prohibit disease from spreading in certain habitats (Hall *et al.*, 2010). However, few lakes have been surveyed for the distribution and abundance of parasites in zooplankton. Thus, we still know very little about the relative importance of various mechanisms that shape the biogeography of disease in freshwater zooplankton. Moreover, we still know little about actual dispersal rates

of either host or parasites. Ongoing theoretical developments in ‘evolving metacommunity’ theory, coupled with other types of network disease models, offer a promising framework in which to integrate ecological and evolutionary mechanisms to study the distribution and abundance of disease (Leibold and Norberg, 2004; Urban and Skelly, 2006; Loeuille and Leibold, 2008; Urban and De Meester, 2009).

DO EPIDEMICS MATTER TO TROPHIC INTERACTIONS AND ECOSYSTEM PROCESSES?

The role that *Daphnia* play in regulating phytoplankton diversity and abundance, nutrient cycling and energy flow is well understood (reviewed in Lampert and Sommer, 2007), but how do parasites alter these interactions? Given that most of these parasites are obligate killers, host densities often decline during epidemics (e.g. Johnson *et al.*, 2006; Hall *et al.*, 2011). Can parasite outbreaks cause or alter a trophic cascade as has been seen in experimental mesocosms (Duffy, 2007)?

If parasite epidemics can have big effects on *Daphnia* populations, can they alter nutrient cycling in lakes? (Frost *et al.*, 2008b) documented significant changes in host stoichiometry as a result of infection by *Pasteuria*. They suggested that infected *Daphnia* may release P at higher rates than their un-infected conspecifics. Could a large enough epidemic influence overall P cycling either through trophic cascades or via altered stoichiometry of host populations? The answer must also consider how infection influences the density and composition of competing grazers, as changes in community structure can influence trophic cascades and nutrient recycling.

Finally, there are a number of unexplored frontiers possible by considering additional parasites or hosts. The role of viruses in freshwater zooplankton is just beginning to be addressed. Natural populations of *Daphnia* harbor circular single-stranded DNA viruses, the prevalence of which can vary throughout the season (Hewson *et al.*, 2013). Will the questions posed above have the same answers for viruses? Will what we have learned from *Daphnia* translate to other species of freshwater zooplankton? Although most studies have focused on *Daphnia* there are also a handful of documented cases of endoparasites of other taxa (e.g. Yan and Larsson, 1988; Thomas *et al.*, 2011). What is needed are more complete natural history studies of parasite ecology and systematics. In many ways we are limited in our ecological and evolutionary studies by a lack of this information. If we are to incorporate parasitism into our understanding of plankton ecology we need more studies on other zooplankton, especially rotifers and copepods.

FUTURE DIRECTIONS

To date, much of the research involving host–parasite interactions has focused either on the ecology of infectious diseases or the evolution of virulence. Given this foundation, the time has come for studies that weave ecology and evolution together, and that integrate processes across levels of biology from physiological defense mechanisms within hosts to life history evolution and clade diversification. The *Daphnia*–parasite system is possibly the best available for studies that will link genome to phenome to understand ecological and evolutionary dynamics of disease. Rapid evolution of both host and parasite is an essential feature of the dynamics of epidemics in *Daphnia* (Duffy and Sivars-Becker, 2007; Wolinska *et al.*, 2008; Wolinska and Spaak, 2009; Duffy *et al.*, 2009; Auld *et al.*, 2012; Duffy *et al.*, 2012). The *P. ramosa*/*Daphnia magna* system is arguably the most intensively studied freshwater zooplankton–parasite system (e.g. Little and Ebert, 1999; Carius *et al.*, 2001; Duncan and Little, 2007; Luijckx *et al.*, 2012). Even here, the genetic structure of natural populations of this parasite remains unknown for most populations, limiting our understanding of the contribution of evolution to these dynamics (Mouton and Ebert, 2008; Andras and Ebert, 2013).

The work of Decaestecker *et al.* (2007, 2013) that takes advantage of the dormant stage of both host and parasite clearly shows that there is evidence of a potential longer-term dynamic. However, one limitation of relying only on what can be reconstructed from the sediments is that it is often impossible to quantify fully both how community structure and seasonal and long-term community dynamics have changed (Jankowski and Straile, 2003). As longer-term data sets accumulate, some of these questions can be resolved. Finally, studies focused on deep time that consider the phylogeographic patterns of both host and parasite are needed. How have those interactions shaped host and parasite life history, both within and among species? How have these phylogenetic differences been modified by adaptation to specific habitats such as lakes versus ponds, or temporal habitat partitioning?

Multi-time scale, joint ecological and evolutionary perspectives are necessary to address a forward looking question: will a warmer world be a sicker world? Mounting evidence indicates that disease prevalence continues to increase among many groups of plant and animal species; furthermore, this increase often ties directly and/or indirectly to global warming (Harvell *et al.*, 1999, 2002; Lafferty *et al.*, 2004; Ward and Lafferty, 2004). Warmer temperatures can both directly enhance parasite fitness and spread by accelerating the thermal physiology of parasites and may also enhance population density, habitat suitability and activity rate (Martens *et al.*, 1999; Harvell

et al., 2002). However, climate change and associated global warming could decrease disease prevalence in some hosts or locations (Hall et al., 2006; Lafferty, 2009; Randolph, 2009). Such outcomes may arise for at least three reasons: changes to habitat suitability (e.g. changes in lake thermal regimes), thermal physiology (host–parasite mis-matches) and changes to the community interactions that influence disease spread. We can start with comparative studies that address these questions in habitats ranging from the tropics to the subtropics to the arctic. What are the latitudinal or other biogeographic patterns of parasitism in freshwater zooplankton?

Zooplankton have long served as model organisms for fundamental research in ecology and evolution. The extensive knowledge of their biology, from genetics to life history and from population to ecosystem dynamics, makes them uniquely valuable in the development and testing of general theory of disease. We suggest that future progress will be made more quickly if emphasis is placed on the integration of processes that govern physiological, ecological and evolutionary aspects of disease.

ACKNOWLEDGEMENTS

Statements in this article are those of the authors and do not necessarily reflect the views of the National Science Foundation. We thank John Megahan, Senior Biological Illustrator, Museum of Zoology, University of Michigan, for Fig. 1. This work was supported by National Science Foundation Grants (DEB 1120804 to C.E.C., DEB 1120316 to S.R.H. and DEB 1305836 to M.A.D.).

REFERENCES

- Altermatt, F. and Ebert, D. (2007) The genotype specific competitive ability does not correlate with infection in natural *Daphnia magna* populations. *Plos One*, **2**, e1280.
- Andras, J. P. and Ebert, D. (2013) A novel approach to parasite population genetics: experimental infection reveals geographic differentiation, recombination and host-mediated population structure in *Pasteuria ramosa*, a bacterial parasite of *Daphnia*. *Mol. Ecol.*, **22**, 972–986.
- Antonovics, J., Boots, M., Ebert, D. et al. (2013) The origin of specificity by means of natural selection: evolved and nonhost resistance in host–pathogen interactions. *Evolution*, **67**, 1–9.
- Auld, S. K. J. R., Hall, S. R. and Duffy, M. A. (2012) Epidemiology of a *Daphnia*-multiparasite system and its implications for the Red Queen. *Plos One*, **7**: e39564.
- Auld, S. K. J. R., Scholenfield, J. A. and Little, T. J. (2010) Genetic variation in the cellular response of *Daphnia magna* (Crustacea: Cladocera) to its bacterial parasite. *Proc. R. Soc. Lond. B*, **277**, 3291–3297.
- Ben-Ami, F., Mouton, L. and Ebert, D. (2008) The effects of multiple infections on the expression and evolution of virulence in a *Daphnia*-endoparasite system. *Evolution*, **62**, 1700–1711.
- Ben-Ami, F., Rigaud, T. and Ebert, D. (2011) The expression of virulence during double infections by different parasites with conflicting host exploitation and transmission strategies. *J. Evol. Biol.*, **24**, 1307–1316.
- Bertram, C. R., Pinkowski, M., Hall, S. R. et al. (2013) Trait-mediated indirect effects, predators, and disease: test of a size-based model. *Oecologia*, **173**, 1023–1032.
- Bieger, A. and Ebert, D. (2009) Expression of parasite virulence at different host population densities under natural conditions. *Oecologia*, **160**, 247–255.
- Bitner, K., Rathhaupt, K. O. and Ebert, D. (2002) Ecological interactions of the microparasite *Caullelya mesnili* and its host *Daphnia galeata*. *Limnol. Oceanogr.*, **47**, 300–305.
- Boersma, M., Spaak, P. and De Meester, L. (1998) Predator-mediated plasticity in morphology, life history, and behavior in *Daphnia*: the uncoupling of responses. *Am. Nat.*, **152**, 237–248.
- Brearley, G., Rhodes, J., Bradley, A. et al. (2012) Wildlife disease prevalence in human-modified landscapes. *Biol. Rev.*, **88**, 427–442.
- Cáceres, C. E. (1997) Temporal variation, dormancy and coexistence: a field test of the storage effect. *Proc. Natl Acad. Sci. USA*, **94**, 9171–9175.
- Cáceres, C. E., Hall, S. R., Duffy, M. A. et al. (2006) Physical structure of lakes constrains epidemics in *Daphnia* populations. *Ecology*, **87**, 1438–1444.
- Cáceres, C. E., Knight, C. J. and Hall, S. R. (2009) Predator-spreaders: predation can enhance parasite success in a planktonic host–parasite system. *Ecology*, **90**, 2850–2858.
- Carius, H. J., Little, T. J. and Ebert, D. (2001) Genetic variation in a host–parasite association: potential for coevolution and frequency-dependent selection. *Evolution*, **55**, 1136–1145.
- Civitello, D. J., Penczykowski, R. M., Hite, J. L. et al. (2013) Potassium stimulates fungal epidemics in a freshwater invertebrate. *Ecology*, **94**, 380–388.
- Daszak, P., Cunningham, A. A. and Hyatt, A. D. (2000) Wildlife ecology - Emerging infectious diseases of wildlife - Threats to biodiversity and human health. *Science*, **287**, 443–449.
- Decaestecker, E., De Gerssem, H., Michalakos, Y. et al. (2013) Damped long-term host–parasite Red Queen coevolutionary dynamics: a reflection of dilution effects? *Ecol. Lett.*, **16**, 1455–1462.
- Decaestecker, E., De Meester, L. and Ebert, D. (2002) In deep trouble: habitat selection constrained by multiple enemies in zooplankton. *Proc. Natl Acad. Sci. USA*, **99**, 5481–5485.
- Decaestecker, E., Gaba, S., Raeymaekers, J. A. M. et al. (2007) Host–parasite ‘Red Queen’ dynamics archived in pond sediment. *Nature*, **450**, 870–873.
- Decaestecker, E., Labbe, P., Ellegaard, K. et al. (2011) Candidate innate immune system gene expression in the ecological model *Daphnia*. *Dev. Comp. Immunol.*, **35**, 1066–1075.
- DeMott, W. R., McKinney, E. N. and Tessier, A. J. (2010) Ontogeny of digestion in *Daphnia*: implications for the effectiveness of algal defenses. *Ecology*, **91**, 540–548.
- Duffy, M. A. (2007) Selective predation, parasitism, and trophic cascades in a bluegill–*Daphnia*–parasite system. *Oecologia*, **153**, 453–460.
- Duffy, M. A., Cáceres, C. E., Hall, S. R. et al. (2010) Temporal, spatial, and between-host comparisons of patterns of parasitism in lake zooplankton. *Ecology*, **91**, 3322–3331.
- Duffy, M. A., Hall, S. R., Cáceres, C. E. et al. (2009) Rapid evolution, seasonality, and the termination of parasite epidemics. *Ecology*, **90**, 1441–1448.

- Duffy, M. A., Hall, S. R., Tessier, A. J. *et al.* (2005) Selective predators and their parasitized prey: are epidemics in zooplankton under top-down control? *Limnol. Oceanogr.*, **50**, 412–420.
- Duffy, M. A., Housley, J. M., Penczykowski, R. M. *et al.* (2011) Unhealthy herds: indirect effects of predators enhance two drivers of disease spread. *Funct. Ecol.*, **25**, 945–953.
- Duffy, M. A., Ochs, J. H., Penczykowski, R. M. *et al.* (2012) Ecological context influences epidemic size and parasite-driven evolution. *Science*, **335**, 1636–1638.
- Duffy, M. A. and Sivers-Becker, L. (2007) Rapid evolution and ecological host-parasite dynamics. *Ecol. Lett.*, **10**, 44–53.
- Duncan, A. B. and Little, T. J. (2007) Parasite-driven genetic change in a natural population of *Daphnia*. *Evolution*, **61**, 796–803.
- Duncan, A. B., Mitchell, S. E. and Little, T. J. (2006) Parasite-mediated selection and the role of sex and diapause in *Daphnia*. *J. Evol. Biol.*, **19**, 1183–1189.
- Ebert, D. (1994) Virulence and local adaptation of a horizontally transmitted parasite. *Science*, **265**, 1084–1086.
- Ebert, D. (2005) *Ecology, Epidemiology, and Evolution of Parasitism in Daphnia* [Internet]. Bethesda, MD: National Library of Medicine (US), National Center for Biotechnology Information. Available from: <http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=Books>.
- Ebert, D. (2008) Host - parasite coevolution: insights from the *Daphnia* - parasite model system. *Curr. Opin. Microbiol.*, **11**, 290–301.
- Ebert, D., Carius, H. J., Little, T. *et al.* (2004) The evolution of virulence when parasites cause host castration and gigantism. *Am. Nat.*, **164**, S19–S32.
- Ebert, D. and Hamilton, W. D. (1996) Sex against virulence: the coevolution of parasitic diseases. *Trends Ecol. Evol.*, **11**, 79–82.
- Freese, H. M. and Schink, B. (2011) Composition and stability of the microbial community inside the digestive tract of the aquatic crustacean *Daphnia magna*. *Microb. Ecol.*, **62**, 882–894.
- Foxman, B. and Goldberg, D. (2010) Why the human microbiome project should motivate epidemiologists to learn ecology. *Epidemiology*, **21**, 757–759.
- Frost, P. C., Ebert, D. and Smith, V. H. (2008a) Responses of a bacterial pathogen to phosphorus limitation of its host. *Ecology*, **89**, 313–318.
- Frost, P. C., Ebert, D. and Smith, V. H. (2008b) Bacterial infection changes the elemental composition of *Daphnia magna*. *J. Anim. Ecol.*, **77**, 1265–1272.
- Hairton, N. G. Jr and Cáceres, C. E. (1996) Distribution of crustacean diapause: micro- and macroevolutionary pattern and process. Symposium on Diapause in the Crustacea: developments in hydrobiology. *Hydrobiologia*, **320**, 27–44.
- Hall, M. D. and Ebert, D. (2012) Disentangling the influence of parasite genotype, host genotype and maternal environment on different stages of bacterial infection in *Daphnia magna*. *Proc. R. Soc. Lond. B*, **279**, 3176–3183.
- Hall, S. R., Becker, C. R., Duffy, M. A. *et al.* (2010) Variation in resource acquisition and use among host clones creates key epidemiological trade-offs. *Am. Nat.*, **176**, 557–565.
- Hall, S. R., Becker, C. R., Duffy, M. A. *et al.* (2011) Epidemic size determines population-level effects of fungal parasites on *Daphnia* hosts. *Oecologia*, **166**, 833–842.
- Hall, S. R., Becker, C. R., Duffy, M. A. *et al.* (2012) A power-efficiency trade-off in resource use alters epidemiological relationships. *Ecology*, **93**, 645–656.
- Hall, S. R., Knight, C. J., Becker, C. R. *et al.* (2009a) Quality matters: resource quality for hosts and the timing of epidemics. *Ecol. Lett.*, **12**, 118–128.
- Hall, S. R., Simonis, J. L., Nisbet, R. M. *et al.* (2009b) Resource ecology of virulence in a planktonic host-parasite system: an explanation using dynamic energy budgets. *Am. Nat.*, **174**, 149–162.
- Hall, S. R., Smyth, R., Becker, C. R. *et al.* (2010) Why Are *Daphnia* in some lakes sicker? Disease ecology, habitat structure, and the plankton. *Bioscience*, **60**, 363–375.
- Hall, S. R., Tessier, A. J., Duffy, M. A. *et al.* (2006) Warmer does not have to mean sicker: temperature and predators can jointly drive timing of epidemics. *Ecology*, **87**, 1684–1695.
- Harvell, C. D., Kim, K., Burkholder, J. M. *et al.* (1999) Review: marine ecology: emerging marine diseases: climate links and anthropogenic factors. *Science*, **285**, 1505–1510.
- Harvell, C. D., Mitchell, C. E., Ward, J. R. *et al.* (2002) Ecology: climate warming and disease risks for terrestrial and marine biota. *Science*, **296**, 2158–2162.
- Havel, J. E., Colbourne, J. K. and Hebert, P. D. N. (2000) Reconstructing the history of intercontinental dispersal in *Daphnia lumholzi* by the use of genetic markers. *Limnol. Oceanogr.*, **45**, 1414–1419.
- Hewson, I., Ng, G., Li, W. *et al.* (2013) Metagenomic identification, seasonal dynamics, and potential transmission mechanisms of a *Daphnia*-associated single-stranded DNA virus in two temperate lakes. *Limnol. Oceanogr.*, **58**, 1605–1620.
- Jankowski, T. and Straile, D. (2003) A comparison of egg-bank and long-term plankton dynamics of two *Daphnia* species, *D. hyalina* and *D. galeata*: Potentials and limits of reconstruction. *Limnol. Oceanogr.*, **48**, 1948–1955.
- Jensen, K. H., Little, T., Skorpung, A. *et al.* (2006) Empirical support for optimal virulence in a castrating parasite. *PLoS Biol.*, **4**, 1265–1269.
- Johnson, P. T. J., Ives, A. R., Lathrop, R. C. *et al.* (2009) Long-term disease dynamics in lakes: causes and consequences of chytrid infections in *Daphnia* populations. *Ecology*, **90**, 132–144.
- Johnson, P. T. J., Longcore, J. E., Stanton, D. E. *et al.* (2006a) Chytrid fungal infections of *Daphnia pulex*: development, ecology, pathology and phylogeny of *Polycaryum laeve*. *Freshwater Biol.*, **51**, 634–648.
- Johnson, P. T. J., Stanton, D. E., Preu, E. R. *et al.* (2006b) Dining on disease: how interactions between infection and environment affect predation risk. *Ecology*, **87**, 1973–1980.
- Killick, S. C., Obbard, D. J., West, S. A. *et al.* (2008) Parasitism and breeding system variation in North American populations of *Daphnia pulex*. *Ecol. Res.*, **23**, 235–240.
- Lafferty, K. D. (2009) Calling for an ecological approach to studying climate change and infectious diseases. *Ecology*, **90**, 932–933.
- Lafferty, K. D., Porter, J. W. and Ford, S. E. (2004) Are diseases increasing in the ocean? *Ann. Rev. Ecol. Evol. System.*, **35**, 31–54.
- Lampert, W. (2011) *Daphnia*: development of a model organism in ecology and evolution. In Kinne, O., ed. *Excellence in Ecology*, Book 21. Olendorf/Luhu: International Ecology Institute.
- Lampert, W. and Sommer, U. (2007) *Limnology: The Ecology of Lakes and Streams*. Oxford University Press.
- Lass, S. and Ebert, D. (2006) Apparent seasonality of parasite dynamics: analysis of cyclic prevalence patterns. *Proc. R. Soc. B Biol. Sci.*, **273**, 199–206.

- Leibold, M. A. (1990) Resources and predation can affect the vertical distribution of zooplankton. *Limnol. Oceanogr.*, **35**, 938–944.
- Leibold, M. A. (1991) Trophic interactions and habitat segregation between competing *Daphnia*. *Oecologia*, **86**, 510–520.
- Leibold, M. A. and Norberg, J. (2004) Biodiversity in metacommunities: plankton as complex adaptive systems?. *Limnol. Oceanogr.*, **49**, 1278–1289.
- Leibold, M. A. and Tessier, A. J. (1991) Contrasting patterns of body size for *Daphnia* species that segregate by habitat. *Oecologia*, **86**, 342–348.
- Little, T. J. and Ebert, D. (1999) Associations between parasitism and host genotype in natural populations of *Daphnia* (Crustacea : Cladocera). *J. Animal Ecol.*, **68**, 134–149.
- Loeuille, N. and Leibold, M. A. (2008) Evolution in metacommunities: on the relative importance of species sorting and monopolization in structuring communities. *Am. Nat.*, **171**, 788–799.
- Luijckx, P., Fienberg, H., Duneau, D. *et al.* (2012) Resistance to a bacterial parasite in the crustacean *Daphnia magna* shows Mendelian segregation with dominance. *Heredity*, **108**, 547–551.
- Martens, P., Kovats, R. S., Nijhof, S. *et al.* (1999) Climate change and future populations at risk from malaria. *Global Environ. Change*, **9**, S89–S107.
- McTaggart, S. J., Conlon, C., Colbourne, J. K. *et al.* (2009) The components of the *Daphnia pulex* immune system as revealed by complete genome sequencing. *BMC Genomics*, **10**, 175.
- Miner, B. E., De Meester, L., Pfrender, M. E. *et al.* (2012) Linking genes to communities and ecosystems: *Daphnia* as an ecogenomic model. *Proc. R. Soc. B Biol. Sci.*, **279**, 1873–1882.
- Mitchell, S. E., Rogers, E. S., Little, T. J. *et al.* (2005) Host-parasite and genotype-by-environment interactions: temperature modifies potential for selection by a sterilizing pathogen. *Evolution*, **59**, 70–80.
- Mouton, L. and Ebert, D. (2008) Variable-number-of-tandem-repeats analysis of genetic diversity in *Pasteuria ramosa*. *Curr. Microbiol.*, **56**, 447–452.
- Mucklow, P. T. and Ebert, D. (2003) Physiology of immunity in the water flea *Daphnia magna*: environmental and genetic aspects of phenoloxidase activity. *Physiol. Biochem. Zool.*, **76**, 836–842.
- Parker, B. J., Spragg, C. J., Altincicek, B. *et al.* (2013) Symbiont-mediated protection against fungal pathogens in pea aphids: a role for pathogen specificity? *Appl. Environ. Microbiol.*, **79**, 2455–2458.
- Pulkkinen, K. and Ebert, D. (2004) Host starvation decreases parasite load and mean host size in experimental populations. *Ecology*, **85**, 823–833.
- Pulkkinen, K. and Ebert, D. (2006) Persistence of host and parasite populations subject to experimental size-selective removal. *Oecologia*, **149**, 72–80.
- Randolph, S. E. (2009) Perspectives on climate change impacts on infectious diseases. *Ecology*, **90**, 927–931.
- Refardt, D. and Ebert, D. (2007) Inference of parasite local adaptation using two different fitness components. *J. Evol. Biol.*, **20**, 921–929.
- Refardt, D. and Ebert, D. (2012) The impact of infection on host competition and its relationship to parasite persistence in a *Daphnia* microparasite system. *Evol. Ecol.*, **26**, 95–107.
- Reissen, H. P. (1999) *Chaoborus* predation and delayed reproduction in *Daphnia*: a demographic modeling approach. *Evol. Ecol.*, **13**, 339–363.
- Stirnadel, H. A. and Ebert, D. (1997) Prevalence, host specificity and impact on host fecundity of microparasites and epibionts in three sympatric *Daphnia* species. *J. Animal Ecol.*, **66**, 212–222.
- Thomas, S. H., Bertram, C., van Rensburg, K. *et al.* (2011) Spatiotemporal dynamics of free-living stages of a bacterial parasite of zooplankton. *Aquatic Microb. Ecol.*, **63**, 265–272.
- Tollrian, R. (1993) Neckteeth formation in *Daphnia pulex* as an example of continuous phenotypic plasticity: morphological effects of *Chaoborus* kairomone concentration and their quantification. *J. Plankton Res.*, **15**, 1309–1318.
- Urban, M. C. and De Meester, L. (2009) Community monopolization: local adaptation enhances priority effects in an evolving metacommunity. *Proc. R. Soc. B*, **276**, 4129–4138.
- Urban, M. C. and Skelly, D. K. (2006) Evolving metacommunities: Toward an evolutionary perspective on metacommunities. *Ecology*, **87**, 1616–1626.
- Ward, J. R. and Lafferty, K. D. (2004) The elusive baseline of marine disease: Are diseases in ocean ecosystems increasing? *Public Library Sci. Biol.*, **2**, 542–547.
- Wilfert, L. and Schmid-Hempel, P. (2008) The genetic architecture of susceptibility to parasites. *BMC Evol. Biol.*, **8**, 187.
- Wolinska, J., Bittner, K., Ebert, D. *et al.* (2006) The coexistence of hybrid and parental *Daphnia*: the role of parasites. *Proc. R. Soc. B Biol. Sci.*, **273**, 1977–1983.
- Wolinska, J., Lively, C. M. and Spaak, P. (2008) Parasites in hybridizing communities: the Red Queen again?. *Trends Parasitol.*, **24**, 121–126.
- Wolinska, J. and Spaak, P. (2009) The cost of being common: evidence from a natural *Daphnia* population. *Evolution*, **63**, 1893–1901.
- Yan, N. D. and Larsson, J. I. R. (1988) Prevalence and inferred effects of microsporidia on *Holopedium gibberum* (Crustacea, Cladocera) in a Canadian Shield lake. *J. Plankton Res.*, **10**, 875–886.
- Yin, M. B., Laforsch, C., Lohr, J. N. *et al.* (2011) Predator-induced defense makes *Daphnia* more vulnerable to parasites. *Evolution*, **65**, 1482–1488.